Reconstruction of phylogenetic trees

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VU medisch centrum

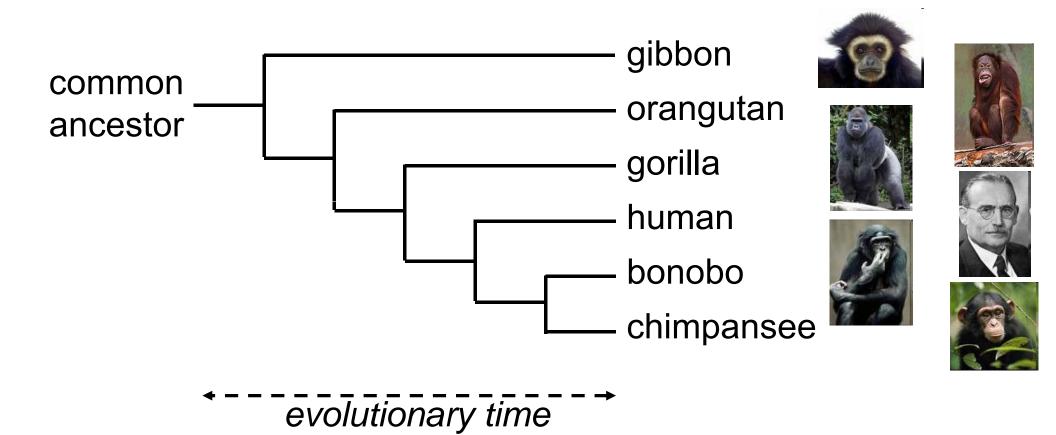


Phylogenetics

"Acceptance of the theory of evolution as the means of explaining observed similarities and differences among organisms invites the construction of trees of descent purporting to show evolutionary relationships"

-- Cavalli-Sforza, Edwards (1967)

Phylogenetics



Phylogenetics is the study of evolutionary relationships between organisms.

Goal

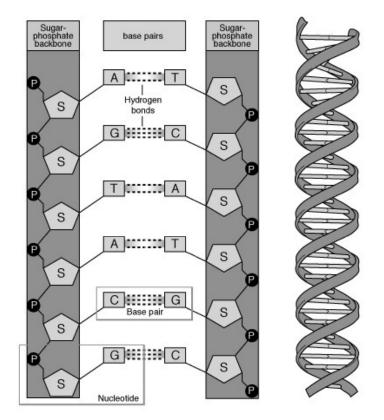
- Reconstruct correct genealogical ties among biogical entities.
- Estimate the time of divergence between organisms.
- Chronicle the sequence of events along evolutionary lineages.

Statistical operationalization: reconstruction of phylogenetic trees on the basis of DNA sequences.

This can also be done on the basis of other characteristics.

Conceptually, **DNA** is an information-carrier.

At the molecular level DNA is a double-stranded polymer existing of four basic molecular units, called *nucleotides*, and denoted by the letters: A, C, G and T.



DNA of each individual is unique, but differences are small: 1 in 500 to 1000 nucleotides differ between two individuals.

Within a population each position in the DNA has a 'predominant' nucleotide.

Over generations this 'pre-dominant'-nucleotide of a position can change by evolution.

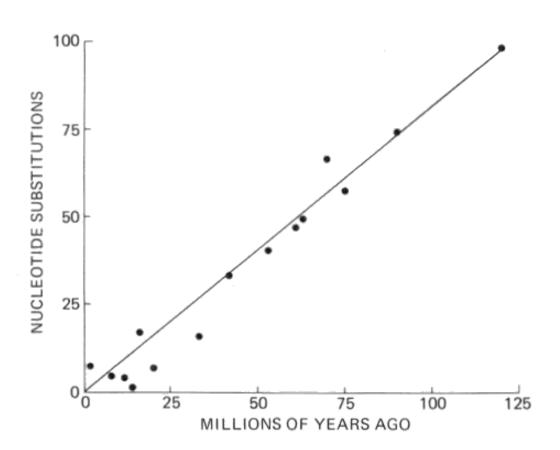
This process is called *substitution*, and takes place over 1000s of generations.

Phylogenetics

Molecular clock-hypothesis

Pair-wise DNA differences between 17 mamal species, plotted against their 'time-ofdivergence', determined from fossil records.

The linear relation suggests that molecular differences between pairs of species are proportional to their 'time-ofdivergence'.

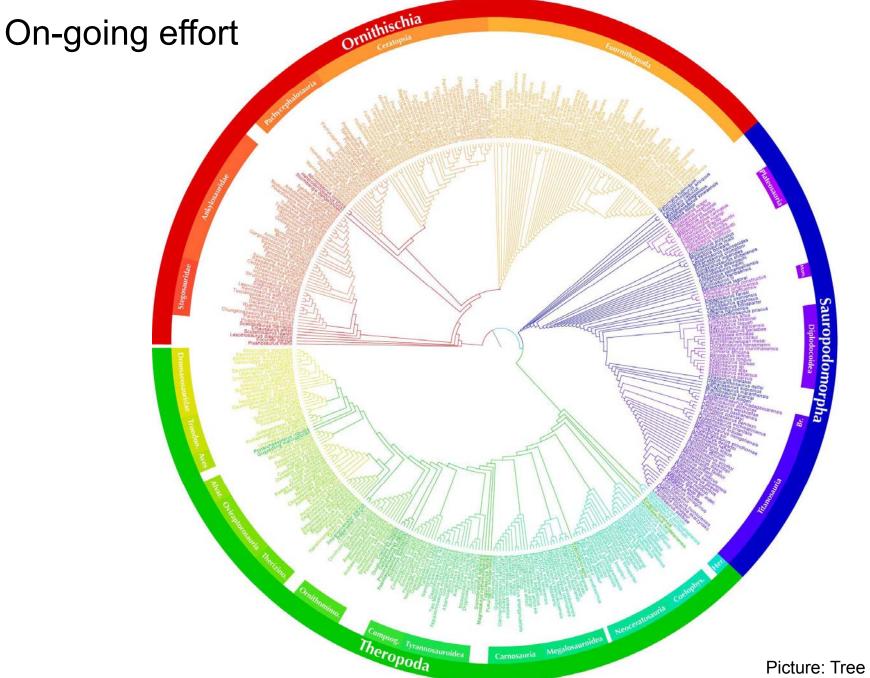


Reconstruction of molecular phylogenetic relations is a step-wise process:

- 1) Select sequences.
- 2) Build a model that describes evolution over time.
- 3) Find the tree that best describes the phylogenetic relations between the sequences.
- 4) Interpret the results.

contribution of the statistician

Phylogenetics



Picture: Tree of Life Web Project

Phylogenetics

The platypus: reptile or mamal?

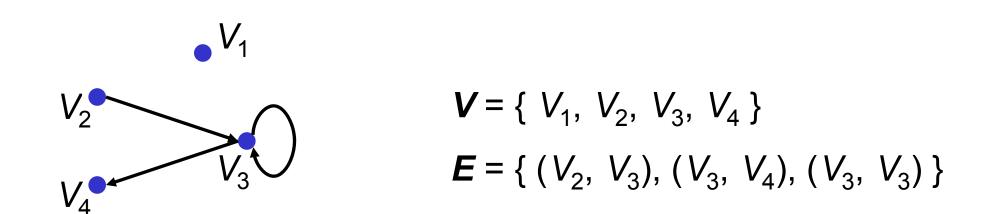
Recently, the genome of the platypus / duck bill has been sequenced.

This revealed: a) +/- 220 My ago separated from the reptiles, b) +/- 170 My ago separated from the mamals, and then evolved separately.

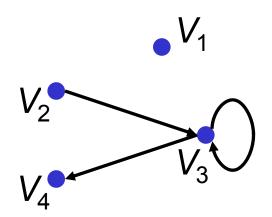
Intermezzo on graphs

A *graph* is a system of connected components. The connections are called *edges*, and components *nodes*.

The *topology* of a graph is a pair (V, E), where V the set of edges and E a subset of $V \times V$.



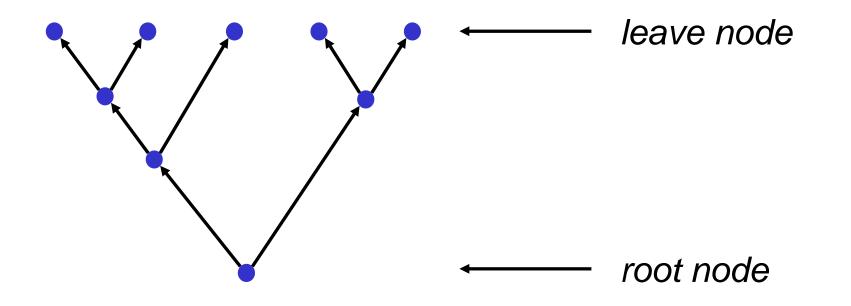
A *path* in a graph is a set of connected edges. In case the begin and end point of a path coincide, the path is called a *cycle*.



Path: (V_2, V_3) , (V_3, V_4) Cycle: (V_3, V_3) If a nodes of a graph are connected (i.e., there is a path between all nodes), the graph is called *connected*.

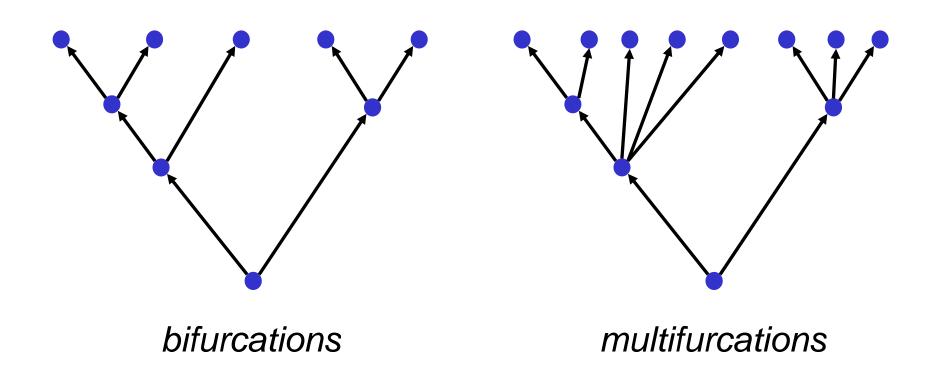
A connected graph that contains no cycles is called a *tree*.

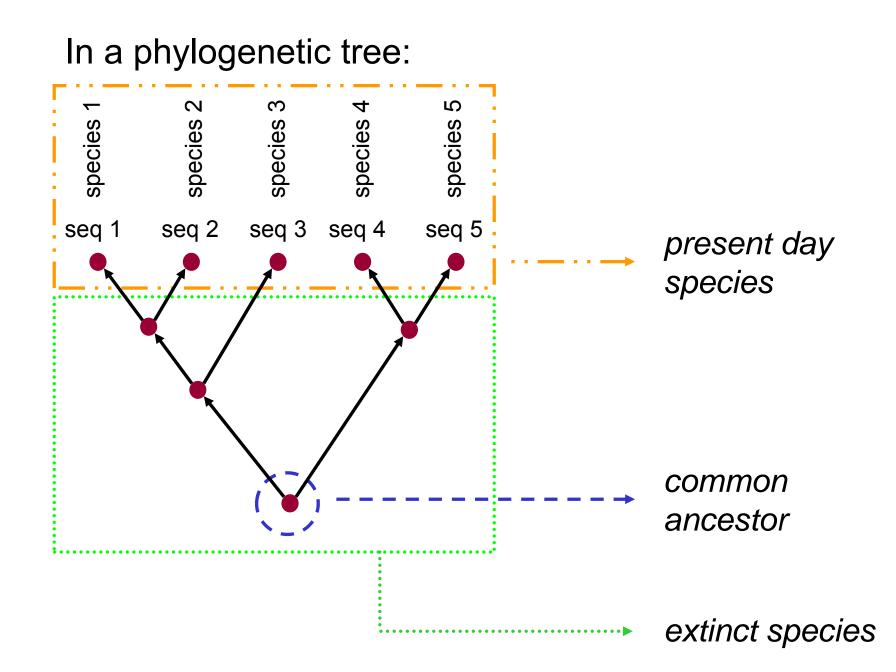
In a *binary tree* every node has either one or three edges, except for the *root node*, if present, that has two edges.



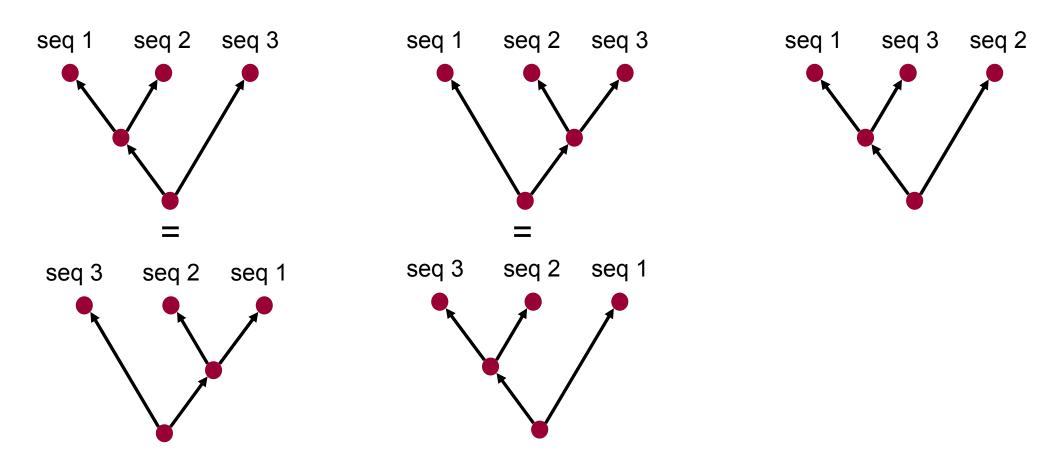
Intermezzo on graphs

Here we only consider binary trees. This rules out the possibility of one species evolving into three or more new species at a particular instance

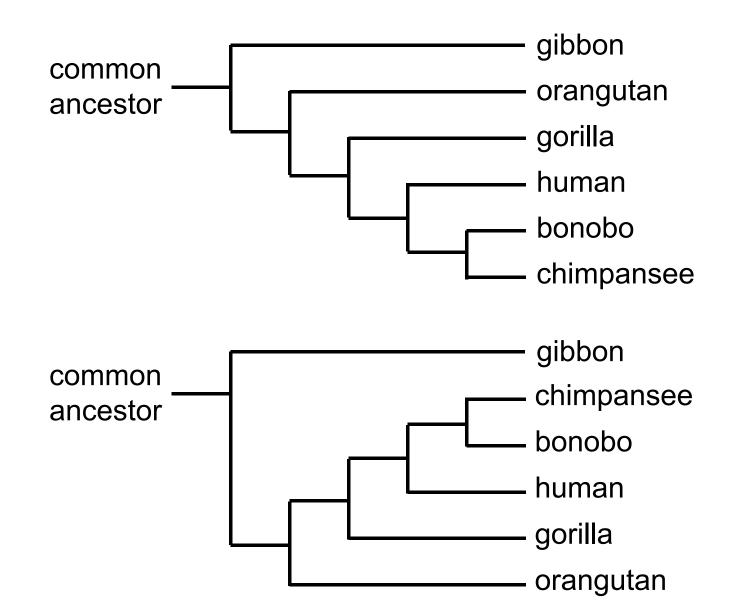




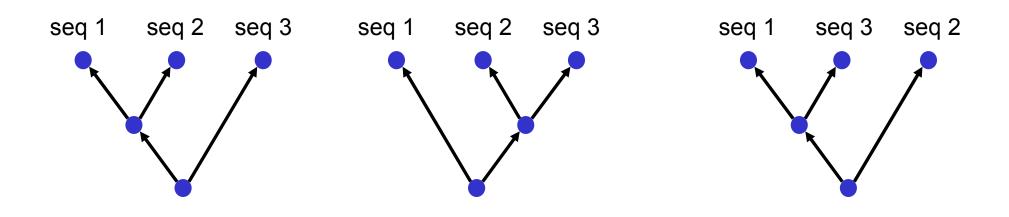
In case of three observed sequences, there are three different trees that connect the sequences:



Hence, the following topologies are equivalent.



If we have three observed sequences, we have three different *rooted* binary trees to connect the three sequences:



The number of possible topologies is enormous. If the number of observed sequences equals n, the number of different *rooted* binary trees is:

(2*n*-3)! / 2^{*n*-2} (*n*-2)!

In case n = 2 : 1 n = 3 : 3 n = 4 : 15 n = 5 : 105.... n = 10 : 34459425

And we have not even considered the branch length!

The number of possible topologies is enormous. If the number of observed sequences equals n, the number of different *unrooted* binary trees is:

(2*n*-5)! / 2^{*n*-3} (*n*-3)!

In case n = 2 : 1 n = 3 : 1 n = 4 : 3 n = 5 : 15 n = 6 : 105.... n = 10 : 2027025n = 11 : 34459425

A model for DNA evolution

For an individual position the substitution process is modeled by a 1st order Markov process with the state space *S*={A, G, C, T}, now grouped by *purines* (A and G) and *pyrimidines* (C and T).

The considered models differ in their parametrization of **P**:

$$\mathbf{P} = \begin{pmatrix} p_{AA} & p_{AG} & p_{AC} & p_{AT} \\ p_{GA} & p_{GG} & p_{GC} & p_{GT} \\ p_{CA} & p_{CG} & p_{CC} & p_{CT} \\ p_{TA} & p_{TG} & p_{TC} & p_{TT} \end{pmatrix}$$

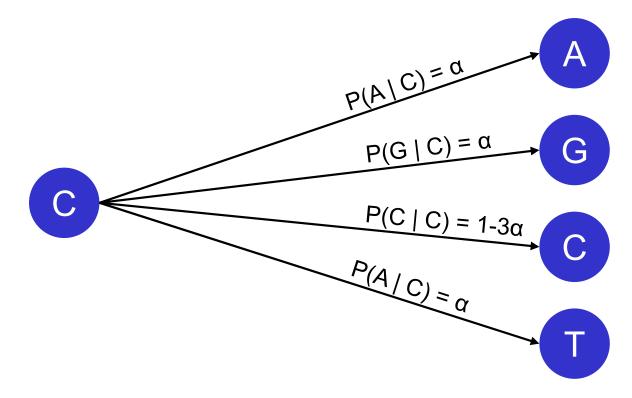
The *Jukes-Cantor model* is a DNA substitution model which assumes that:

- each base in the sequence has an equal probability of being substituted.
- if a nucleotide substitution occurs, all other nucleotides have the same probability to replace it.

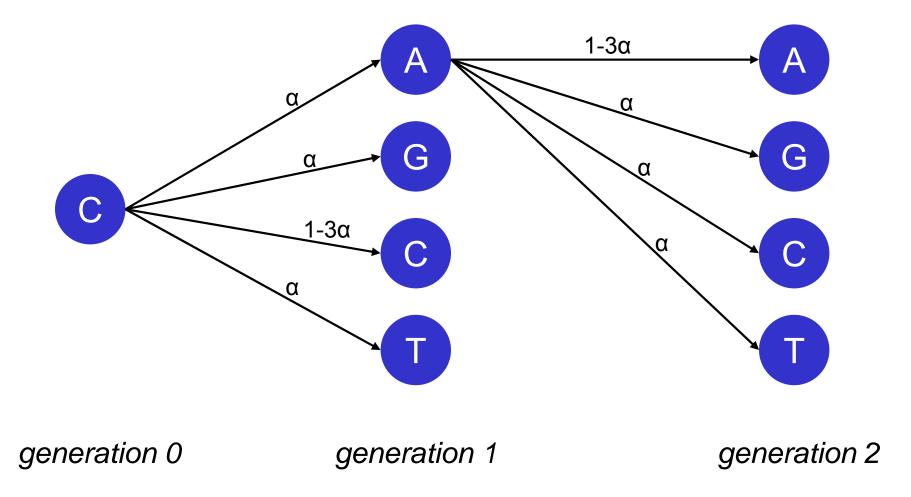
(As a results we expect an equal frequency of the four bases in the resulting DNA sequence.)

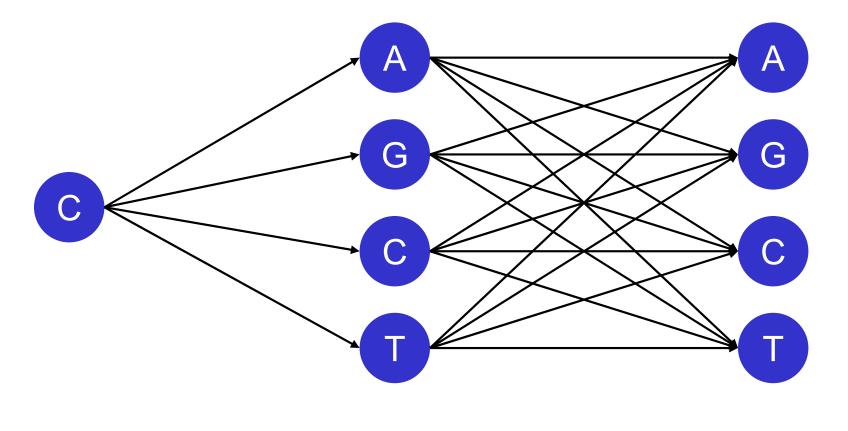
We have:

- probability α of C to substitute by A,
- probability α of C to substitute by G,
- probability α of C to substitute by T,
- probability 1-3 α of C not to substitute.



generation 0

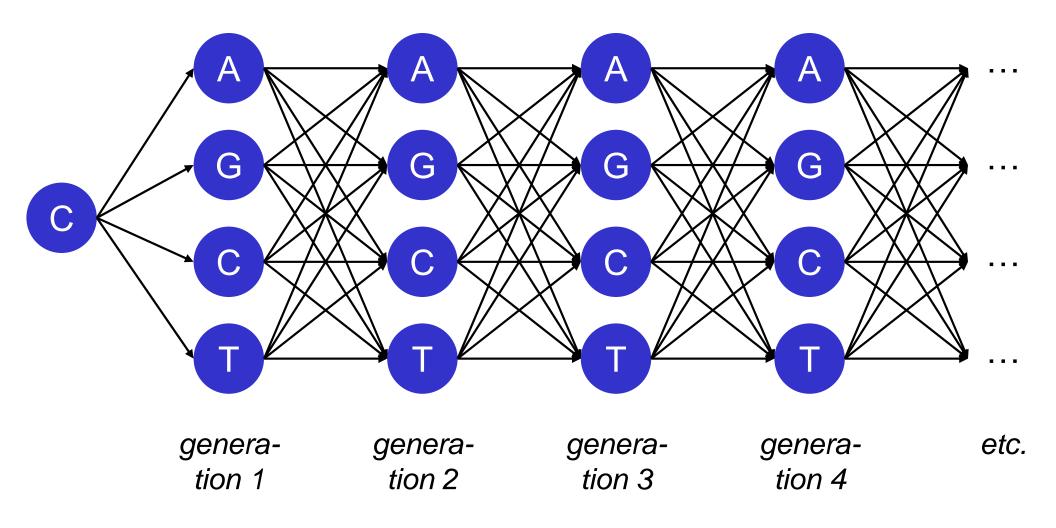




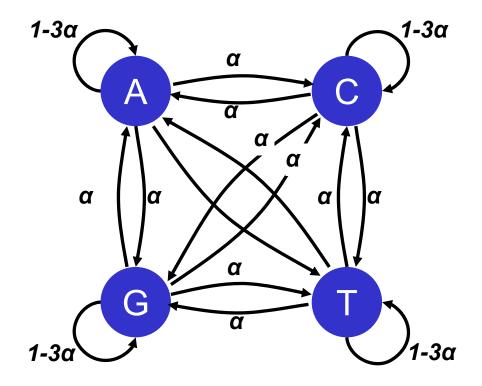
generation 0

generation 1

generation 2



Over 1000s of generations (time homogeneity):

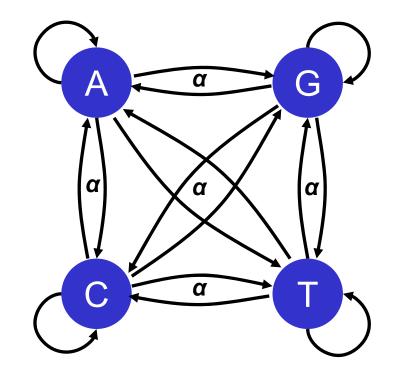


The Jukes-Cantor transition matrix:

$$\mathbf{P} = \begin{pmatrix} 1-3\alpha & \alpha & \alpha & \alpha \\ \alpha & 1-3\alpha & \alpha & \alpha \\ \alpha & \alpha & 1-3\alpha & \alpha \\ \alpha & \alpha & \alpha & 1-3\alpha \end{pmatrix}$$

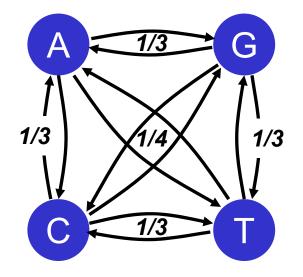
where

- $-\alpha < \frac{1}{3}$,
- α depends on the step size.



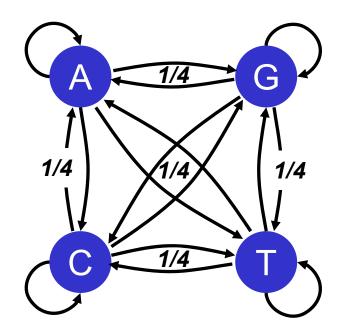
Always substitute if
$$\alpha = 1/3$$
:

$$\mathbf{P} = \begin{pmatrix} 0 & 1/3 & 1/3 & 1/3 \\ 1/3 & 0 & 1/3 & 1/3 \\ 1/3 & 1/3 & 0 & 1/3 \\ 1/3 & 1/3 & 1/3 & 0 \end{pmatrix}$$



No Markov property if $\alpha = 1/4$:

$$\mathbf{P} = \begin{pmatrix} 1/4 & 1/4 & 1/4 & 1/4 \\ 1/4 & 1/4 & 1/4 & 1/4 \\ 1/4 & 1/4 & 1/4 & 1/4 \\ 1/4 & 1/4 & 1/4 & 1/4 \end{pmatrix}$$



Properties of the Jukes-Cantor model

The eigenvalues of **P**:

$$λ = 1$$
, 1-4α, 1-4α, 1-4α.

The stationary distribution corresponding to λ =1:

$$\mathbf{\phi} = (\frac{1}{4}, \frac{1}{4}, \frac{1}{4}, \frac{1}{4})^{\mathsf{T}}$$

Indeed, after enough generations all four states are equally likely. That is, all four nucleotides are equally likely to be the predominant nucleotide at the position under consideration.

Properties of the Jukes-Cantor model

Its spectral decomposition:

Properties of the Jukes-Cantor model

Consider a stationary 1st order Markov chain with a Jukes-Cantor transition matrix. The probability of no substitution is given by:

$$P(X_t = A, X_{t-1} = A, ..., X_0 = A) = P(A | A)^t P(A)$$

= (1-3\alpha)^t \varphi_A
= (1-3\alpha)^t / 4

Given that X_0 =A, the probability that A will be the predominant nucleotide at time *t* is given by:

$$\frac{1}{4} + \frac{3}{4} (1-4\alpha)^{t}$$

Properties of the Jukes-Cantor model

Now we know **P** and ϕ , and, hence, we can assess the reversibility of the Jukes-Cantor model by means of checking the detailed balance equations:

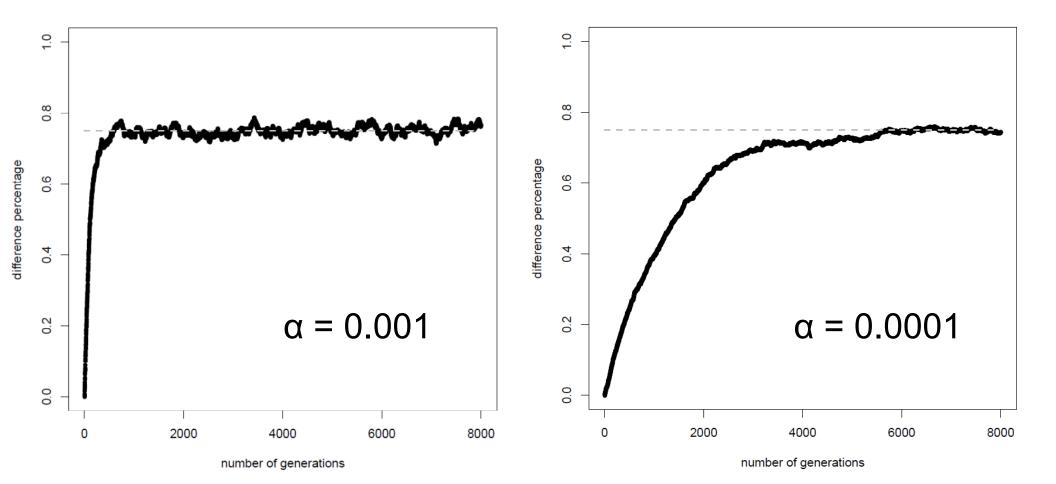
 $\varphi_i p_{ij} = \varphi_j p_{ji}$ for all i and j.

Recall

In order for the Jukes-Cantor model to link one species to another (via a common ancestor), the transition matrix **P** needs to be reversible.

Models for DNA evolution (JC69)

Proportion of site differences between two sequences in the Jukes-Canter model plotted against time (# generations), starting from the common ancestor.



The *Kimura model* is a generalization of the Jukes-Cantor model. It allows for different transition and transversion probabilities.

Similar to the Jukes-Cantor model, the Kimura is symmetrical. Therefore, after enough time it is equally likely for a base to be a purine or a pyrimidine.

Within the purine and pyrimidine categories there is complete symmetry between the nucleotides.

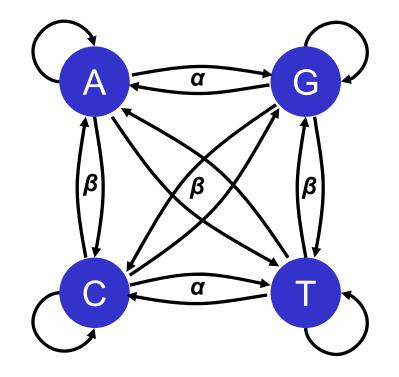
The Kimura transition matrix:

$$\mathbf{P} = \begin{pmatrix} 1 - \alpha - 2\beta & \alpha & \beta & \beta \\ \alpha & 1 - \alpha - 2\beta & \beta & \beta \\ \beta & \beta & 1 - \alpha - 2\beta & \alpha \\ \beta & \beta & \alpha & 1 - \alpha - 2\beta \end{pmatrix}$$

where

- $-\alpha+2\beta<1,$
- α, β depend on the step size.

Take $\alpha = \beta$: Jukes-Cantor.



Properties of the Kimura model

The eigenvalues of **P**:

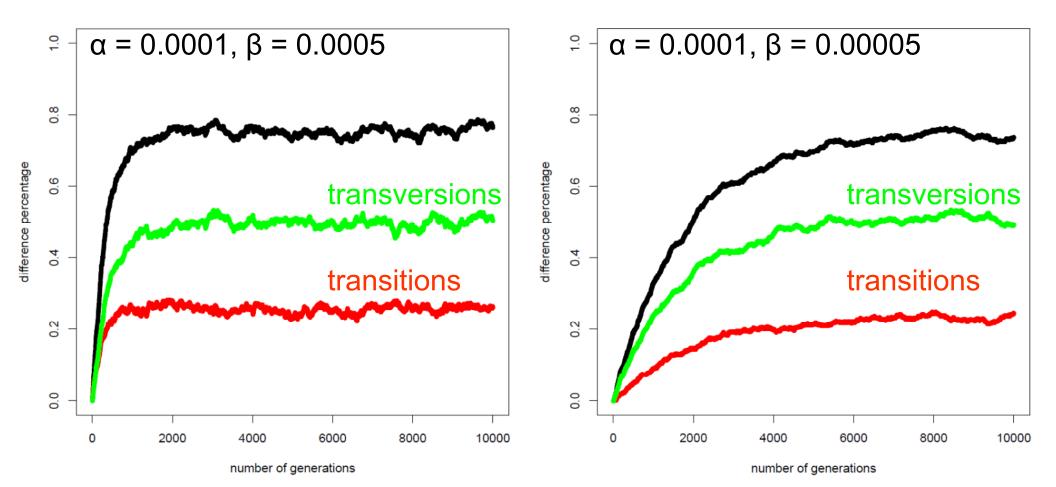
$$λ = 1$$
, 1-4β, 1-2(α+β), 1-2(α+β).

The stationary distribution corresponding to λ =1:

 $\mathbf{\phi} = (\frac{1}{4}, \frac{1}{4}, \frac{1}{4}, \frac{1}{4})^{\mathsf{T}}$

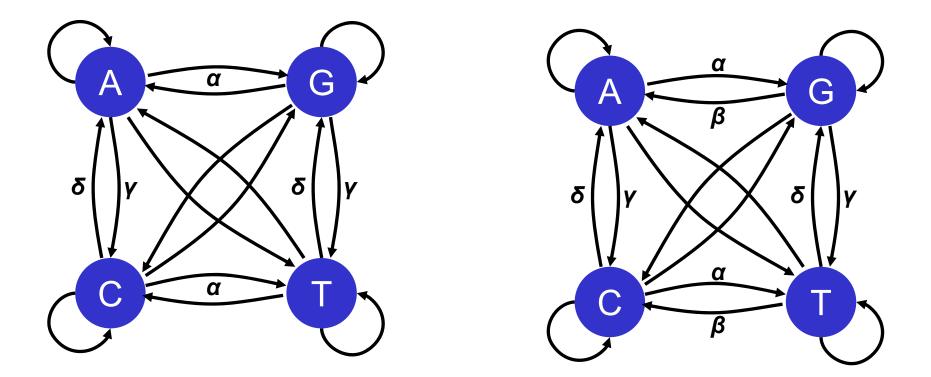
The Kimura model is reversible (**P** is symmetric and $\boldsymbol{\phi}$ uniform).

Proportion of site differences between two sequences in the Kimura model plotted against time (# generations), starting from the common ancestor.



The Kimura model has been generalized to allow, e.g.:

- The transition probability to differ from the transversion probability.
- Different within-transition and within-transversion substitution probabilities.



The *Felsenstein model* is also a generalization of the Jukes-Cantor model. It relaxes the (implicit) assumption of the JC and Kimura model, both having a uniform stationary distribution.

In the Felsenstein model the probability of substitution of any nucleotide by another is proportional to the stationary probability of the substituting nucleotide.

The Felsenstein model does not distinguish between purines and pyrimidines.

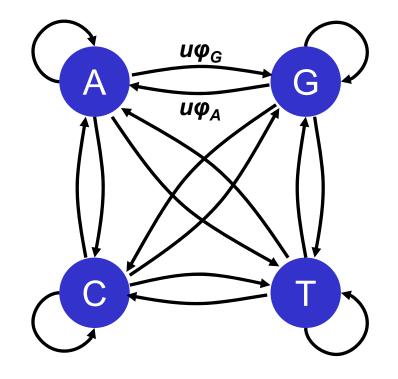
The Felsenstein transition matrix:

$$\mathbf{P} = \begin{pmatrix} 1 - u + u \varphi_A & u \varphi_G & u \varphi_C & u \varphi_T \\ u \varphi_A & 1 - u + u \varphi_G & u \varphi_C & u \varphi_T \\ u \varphi_A & u \varphi_G & 1 - u + u \varphi_C & u \varphi_T \\ u \varphi_A & u \varphi_G & u \varphi_C & 1 - u + u \varphi_T \end{pmatrix}$$

where

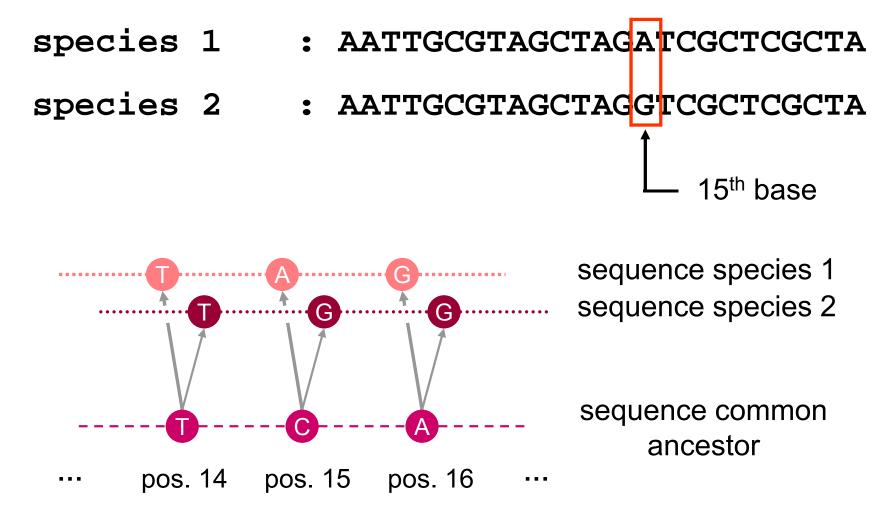
- $\phi_{A} + \phi_{G} + \phi_{C} + \phi_{T} = 1.$
- *u* a model parameter.

Take $\phi_A = \phi_G = \phi_C = \phi_T = \frac{1}{4}$: Jukes-Cantor.



The likelihood: a simple example

Consider two homologous sequences sampled from two different species (with a common ancestor):



What is the likelihood of observing these two sequences?

Let

- X denote the sequence data of both species, and
- X_{ij} denote the nucleotide at position *j*=1, ...,25 of species *i*.

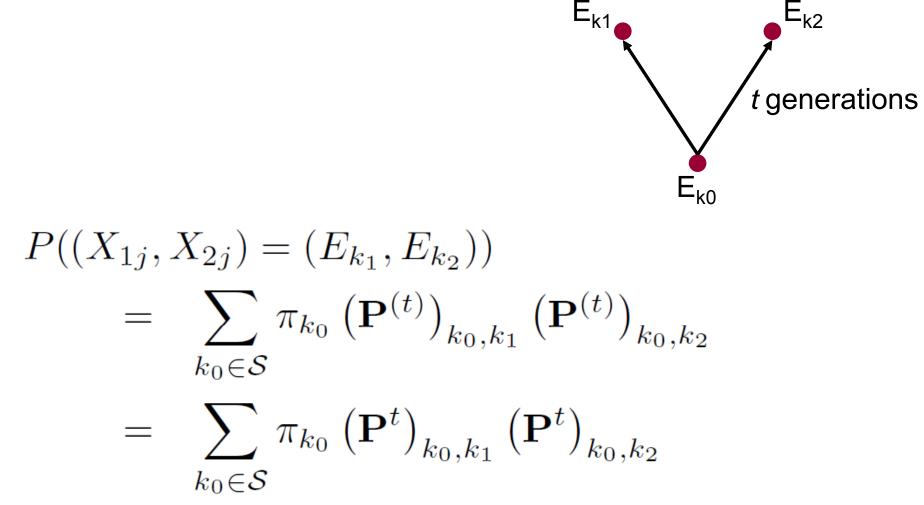
The likelihood for the Jukes-Cantor model is then:

 $L(\mathbf{X}) = P(\mathbf{X})$

which, assuming sites evolve independently, factorizes to

$$= \prod_{j=1}^{25} P((X_{1j}, X_{2j}))$$

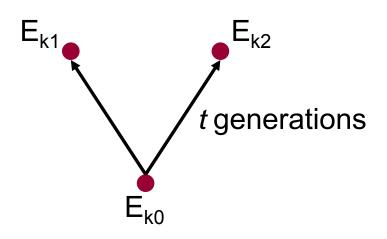
Assuming $(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})$ and that the species have evolved separately *t* generations since the common ancestor, then:



Note

The life time of a generation may differ between the two present day organisms. In particular, if an evolutionary long time has passed since the common ancestor.

The solution is to use the actual time passed since the common ancestor. Modeling this requires continuous time Markov chains. Not treated here.



In order to write down the likelihood, recall - The Chapman-Kolmogorov equations:

$$p_{k_1k_2}^{(t_1+t_2)} = \sum_{k_0 \in \mathcal{S}} p_{k_1k_0}^{(t_1)} p_{k_0k_2}^{(t_2)}$$

- The reversibility of the Jukes-Cantor model:

$$\pi_{k_1} \, p_{k_1 k_2} \quad = \quad \pi_{k_2} \, p_{k_2 k_1}$$

- The symmetry of the JC transition matrix P.
- Combining the last two yields:

$$\pi_{k_1} p_{k_1 k_2}^{(t)} = \pi_{k_2} p_{k_2 k_1}^{(t)}$$

$$L(\mathbf{X}) = \prod_{j=1}^{25} P((X_{1j}, X_{2j}))$$

=
$$\prod_{j=1}^{25} \prod_{k_1, k_2 \in \mathcal{S}} \left[P((X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})) \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

$$L(\mathbf{X}) = \prod_{j=1}^{25} P((X_{1j}, X_{2j}))$$

=
$$\prod_{j=1}^{25} \prod_{k_1, k_2 \in \mathcal{S}} \left[P((X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})) \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

=
$$\prod_{j=1}^{25} \prod_{k_1, k_2 \in \mathcal{S}} \left[\sum_{k_0 \in \mathcal{S}} \pi_{k_0} \left(\mathbf{P}^t \right)_{k_0, k_1} \left(\mathbf{P}^t \right)_{k_0, k_2} \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

- 1) substitute previously derived expression for probability of individual observation
- 2) substitution rates are the same for all sites

$$L(\mathbf{X}) = \prod_{j=1}^{25} P((X_{1j}, X_{2j}))$$

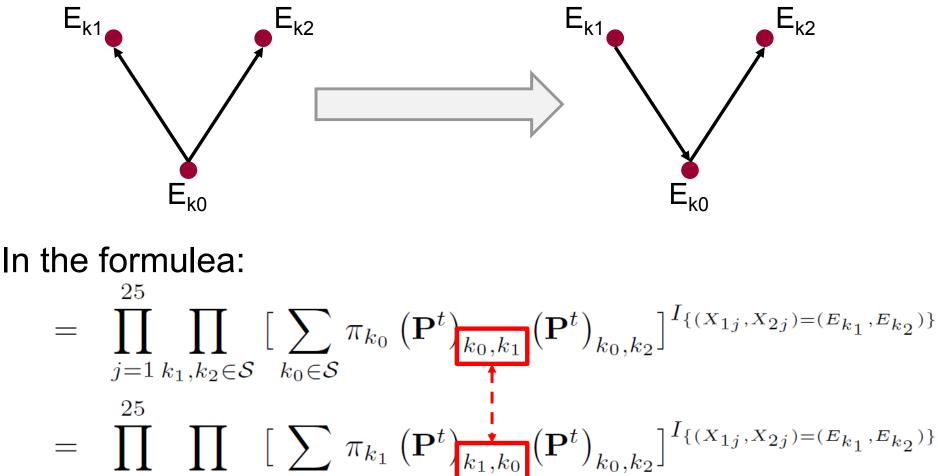
$$= \prod_{j=1}^{25} \prod_{k_1, k_2 \in S} \left[P((X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})) \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

$$= \prod_{j=1}^{25} \prod_{k_1, k_2 \in S} \left[\sum_{k_0 \in S} \pi_{k_0} \left(\mathbf{P}^t \right)_{k_0, k_1} \left(\mathbf{P}^t \right)_{k_0, k_2} \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

$$= \prod_{j=1}^{25} \prod_{k_1, k_2 \in S} \left[\sum_{k_0 \in S} \pi_{k_1} \left(\mathbf{P}^t \right)_{k_1, k_0} \left(\mathbf{P}^t \right)_{k_0, k_2} \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

use the time reversibility of the JC model

By using the time reversibility of the JC model, we have reversed one arrow of the phylogenetic tree:



i=1 $k_1,k_2 \in S$ $k_0 \in S$

25 $L(\mathbf{X}) = \prod P((X_{1j}, X_{2j}))$ j=125 $= \prod \left[P((X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})) \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$ j=1 $k_1,k_2 \in \mathcal{S}$ 25 $= \prod \left[\sum \pi_{k_0} \left(\mathbf{P}^t \right)_{k_0, k_1} \left(\mathbf{P}^t \right)_{k_0, k_2} \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$ j=1 k_1 , $k_2 \in S$ $k_0 \in S$ 25 $= \prod \left[\sum \pi_{k_1} \left(\mathbf{P}^t \right)_{k_1, k_0} \left(\mathbf{P}^t \right)_{k_0, k_2} \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$ $j=1 k_1, k_2 \in \mathcal{S} \quad k_0 \in \mathcal{S}$ 25 $= \prod_{j=1}^{I} \prod_{k_1,k_2 \in \mathcal{S}} \left[\pi_{k_1} \sum_{k_0 \in \mathcal{S}} \left(\mathbf{P}^t \right)_{k_1,k_0} \left(\mathbf{P}^t \right)_{k_0,k_2} \right]^{I_{\{(X_{1j},X_{2j})=(E_{k_1},E_{k_2})\}}}$

bringing π_{k1} outside the sum

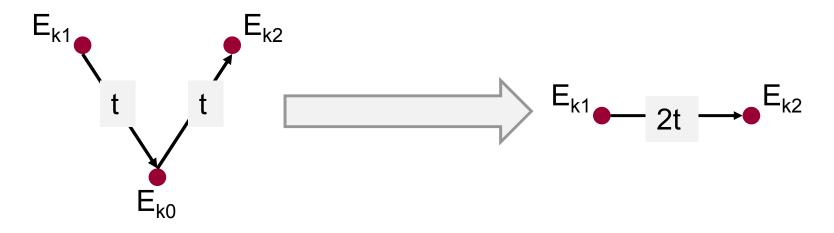
$$L(\mathbf{X}) = \prod_{j=1}^{25} P((X_{1j}, X_{2j}))$$

=
$$\prod_{j=1}^{25} \prod_{k_1, k_2 \in \mathcal{S}} \left[P((X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})) \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

use the Chapman-Kolmogorov equations

$$= \prod_{j=1}^{25} \prod_{k_1,k_2 \in \mathcal{S}} \left[\pi_{k_1} \sum_{k_0 \in \mathcal{S}} \left(\mathbf{P}^t \right)_{k_1,k_0} \left(\mathbf{P}^t \right)_{k_0,k_2} \right]^{I_{\{(X_{1j},X_{2j})=(E_{k_1},E_{k_2})\}}} \\ = \prod_{j=1}^{25} \prod_{k_1,k_2 \in \mathcal{S}} \left[\pi_{k_1} \left(\mathbf{P}^{2t} \right)_{k_1,k_2} \right]^{I_{\{(X_{1j},X_{2j})=(E_{k_1},E_{k_2})\}}}$$

By using the Chapman-Kolmogorov equations, we removed the common ancestor from the phylogenetic tree:



In the formulea:

$$= \prod_{j=1}^{25} \prod_{k_1,k_2 \in \mathcal{S}} \left[\pi_{k_1} \sum_{k_0 \in \mathcal{S}} \left(\mathbf{P}^t \right)_{k_1,k_0} \left(\mathbf{P}^t \right)_{k_0,k_2} \right]^{I_{\{(X_{1j},X_{2j})=(E_{k_1},E_{k_2})\}}} \\ = \prod_{j=1}^{25} \prod_{k_1,k_2 \in \mathcal{S}} \left[\pi_{k_1} \left(\mathbf{P}^{2t} \right)_{k_1,k_2} \right]^{I_{\{(X_{1j},X_{2j})=(E_{k_1},E_{k_2})\}}}$$

The likelihood can be further simplified, when exploiting the spectral decomposition the JC *t*-step transition matrix:

Finally, we have:

$$L(\mathbf{X}) = \prod_{j=1}^{25} \prod_{k_1, k_2 \in S} \left[\pi_{k_1} \left(\mathbf{P}^{2t} \right)_{k_1, k_2} \right]^{I_{\{(X_{1j}, X_{2j}) = (E_{k_1}, E_{k_2})\}}}$$

$$= \prod_{j=1}^{25} \frac{1}{4} \left[\frac{1}{4} + \frac{3}{4} (1 - 4\alpha)^{2t} \right]^{I_{\{X_{1j} = X_{2j}\}}}$$

$$\times \left[\frac{1}{4} - \frac{1}{4} (1 - 4\alpha)^{2t} \right]^{I_{\{X_{1j} \neq X_{2j}\}}}$$

where we have used that the stationary distribution of the JC model is uniform.

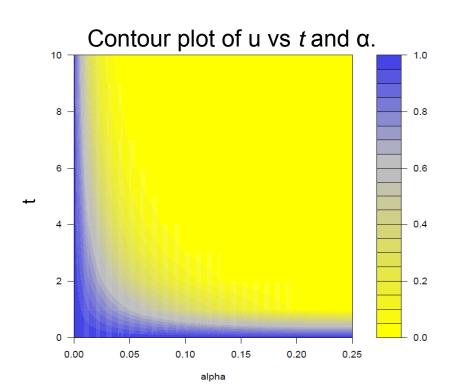
From the likelihood, it is clear either *t* or α is identifiable not both. Many combinations (α , *t*) yield the same likelihood.

In the absence of external evidence of α , we replace:

$$u = (1 - 4\alpha)^{2t}$$

and obtain:

$$L(\mathbf{X}) = \prod_{j=1}^{25} \frac{1}{4} \left[\frac{1}{4} + \frac{3}{4} u \right]^{I_{\{X_{1j}=X_{2j}\}}} \left[\frac{1}{4} - \frac{1}{4} u \right]^{I_{\{X_{1j}\neq X_{2j}\}}}$$



To estimate *u*, maximize the log-likehood:

$$\log\left(\frac{1}{4} + \frac{3}{4}u\right)\sum_{j=1}^{25} I_{\{X_{1j}=X_{2j}\}} + \log\left(\frac{1}{4} - \frac{1}{4}u\right)\sum_{j=1}^{25} I_{\{X_{1j}\neq X_{2j}\}}$$

This yields:

$$\hat{u} = \frac{3 \sum_{j=1}^{25} I_{\{X_{1j}=X_{2j}\}} - \sum_{j=1}^{25} I_{\{X_{1j}\neq X_{2j}\}}}{3 \sum_{j=1}^{25} I_{\{X_{1j}=X_{2j}\}} + 3 \sum_{j=1}^{25} I_{\{X_{1j}\neq X_{2j}\}}}$$

Check that this is indeed a maximum.

For our two-species example, with sequences:

- species 1 : AATTGCGTAGCTAGATCGCTCGCTA
- species 2 : AATTGCGTAGCTAGGTCGCTCGCTA

the ML estimate equals:

$$\hat{u} = \frac{3 \times 24 - 1}{3 \times 24 + 3 \times 1} = \frac{71}{75}$$

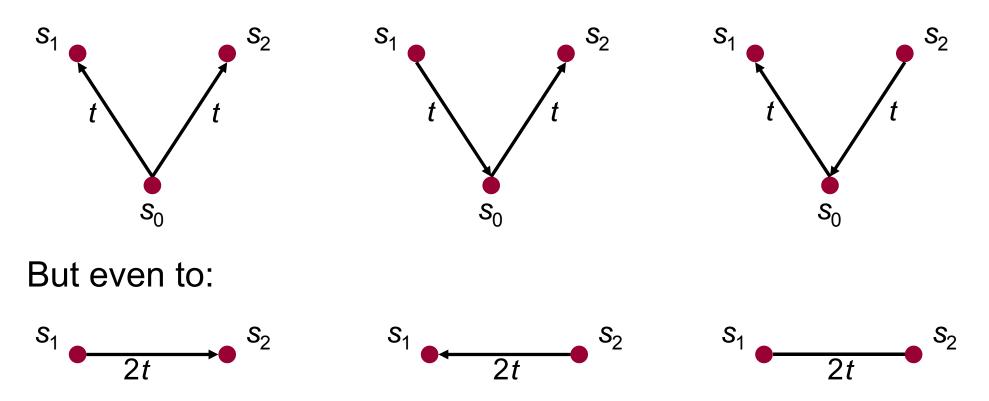
Assuming the substitution rate (α) is 1 in a million, we get:

$$\hat{t} = \log(71/75) / \log(1 - \alpha/4) \approx 219233$$

This estimates suggests that the two species shared a common ancestor 219233 generations ago.

The pulley principle

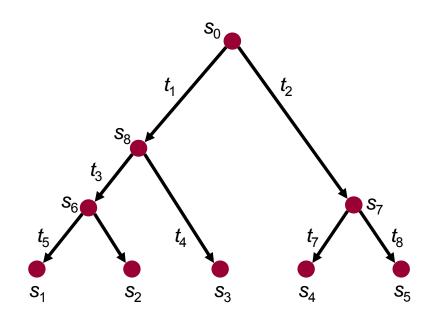
Due to reversibility, likelihood of trees below are equivalent:

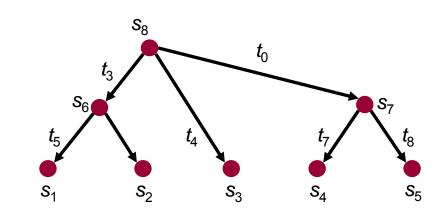


Pulley principle

The root node may be moved to any of the nodes without changing the likelihood.

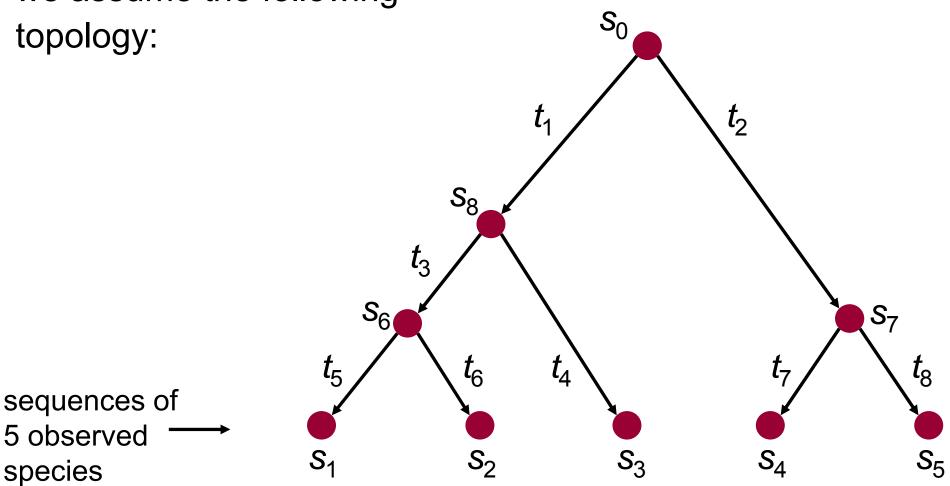
Due to the pulley principle, the likelihood of the following trees is equivalent:





Consider the case where:

- DNA sequences from (say) 5 species are available.
- the sequences consist of (say) 25 bases.
- we assume the following topology:



Step 1

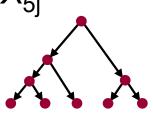
Assume the 25 sites evolve independently. The probability of evolution from (say) node / species s_7 to s_5 then becomes:

$$P(\mathbf{X}_7 \xrightarrow{t_8} \mathbf{X}_5) = \prod_{j=1}^{25} P(X_{7j} \xrightarrow{t_8} X_{5j})$$

where

$$P(X_{7j} \xrightarrow{t_8} X_{5j})$$

denotes the (conditional) probability of X_{7j} evolving to X_{5j} in t_8 generations.



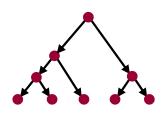
Step 1

Recall: the probability of the nucleotide at site *j* changing from X_{7j} in sequence 7 to X_{5j} in sequence 5 in t_8 generations, denoted by:

$$P(X_{7j} \xrightarrow{t_8} X_{5j})$$

is given by a multiple of the transition matrix of the evolutionary model of choice. Hence,

$$P(X_{7j} \xrightarrow{t_8} X_{5j}) = (\mathbf{P}^{t_8})_{X_{7j}, X_{5j}}$$



Step 2

If the sequence of all nodes / species $(s_0, ..., s_8)$ are known, the likelihood is given by:

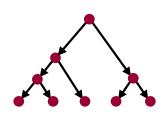
$$L = P(\mathbf{X}_{0})$$

$$P(\mathbf{X}_{0} \xrightarrow{t_{2}} \mathbf{X}_{7}) P(\mathbf{X}_{0} \xrightarrow{t_{1}} \mathbf{X}_{8})$$

$$P(\mathbf{X}_{7} \xrightarrow{t_{7}} \mathbf{X}_{4}) P(\mathbf{X}_{7} \xrightarrow{t_{8}} \mathbf{X}_{5})$$

$$P(\mathbf{X}_{8} \xrightarrow{t_{4}} \mathbf{X}_{3}) P(\mathbf{X}_{8} \xrightarrow{t_{3}} \mathbf{X}_{6})$$

$$P(\mathbf{X}_{6} \xrightarrow{t_{5}} \mathbf{X}_{1}) P(\mathbf{X}_{6} \xrightarrow{t_{6}} \mathbf{X}_{2})$$



Step 3

Since only the sequences of nodes $n_1, ..., n_5$ are observed, the likelihood has to be summed over all possible sequences for the unobserved nodes:

 $L = \sum \sum \sum \sum \sum [P(\mathbf{X}_0)]$ $\mathbf{X}_0 \ \mathbf{X}_6 \ \mathbf{X}_7 \ \mathbf{X}_8$ $P(\mathbf{X}_0 \xrightarrow{t_2} \mathbf{X}_7) P(\mathbf{X}_0 \xrightarrow{t_1} \mathbf{X}_8)$ $P(\mathbf{X}_7 \xrightarrow{t_7} \mathbf{X}_4) P(\mathbf{X}_7 \xrightarrow{t_8} \mathbf{X}_5)$ $P(\mathbf{X}_8 \xrightarrow{t_4} \mathbf{X}_3) P(\mathbf{X}_8 \xrightarrow{t_3} \mathbf{X}_6)$ $P(\mathbf{X}_6 \xrightarrow{t_5} \mathbf{X}_1) P(\mathbf{X}_6 \xrightarrow{t_6} \mathbf{X}_2)]$

Step 3 (computational efficiency)

This likelihood can be calculated by exploiting the conditional likelihoods, e.g.:

$$L((X_{4j}, X_{5j}) | X_{7j}) = \left[\sum_{X_{4j}} P(X_{7j} \xrightarrow{t_7} X_{4j}) L(X_{4j}) \right] \left[\sum_{X_{5j}} P(X_{7j} \xrightarrow{t_8} X_{5j}) L(X_{5j}) \right]$$

which yields:

$$L = \sum_{\mathbf{X}_{0}} P(\mathbf{X}_{0}) \left[\left[\sum_{\mathbf{X}_{7}} P(\mathbf{X}_{0} \xrightarrow{t_{2}} \mathbf{X}_{7}) P(\mathbf{X}_{7} \xrightarrow{t_{7}} \mathbf{X}_{4}) P(\mathbf{X}_{7} \xrightarrow{t_{8}} \mathbf{X}_{5}) \right] \\ \times \left[\sum_{\mathbf{X}_{8}} P(\mathbf{X}_{0} \xrightarrow{t_{1}} \mathbf{X}_{8}) P(\mathbf{X}_{8} \xrightarrow{t_{4}} \mathbf{X}_{3}) P(\mathbf{X}_{8} \xrightarrow{t_{3}} \mathbf{X}_{6}) \\ \times \left[\sum_{\mathbf{X}_{6}} P(\mathbf{X}_{6} \xrightarrow{t_{5}} \mathbf{X}_{1}) P(\mathbf{X}_{6} \xrightarrow{t_{6}} \mathbf{X}_{2}) \right] \right] \right]$$

Step 3 (computational efficiency)

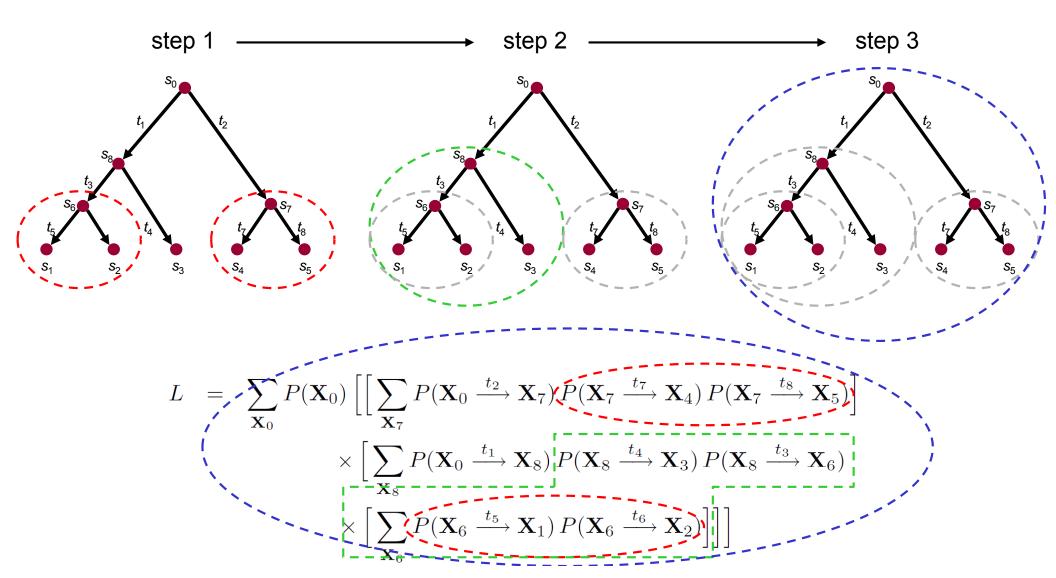
Without the exploitation of the conditional likelihood, calculation of the likelihood required the evaluation of 4⁴=256 combinations (4 hidden nodes, 4 nucleotides).

In the reformulation on the previous slide, the likelihood is evaluated in for 4 * (4+4+4) = 48 steps.

This is (approximately) a factor 5!!!

The likelihood: another example

Pruning: calculate the likelihood by proceeding from the leaves towards the root.



The likelihood: another example

Step 4

As also the topology is in fact unobserved, we need to sum the likelihood from the previous step over all possible topologies.

The pulley principle comes to the rescue, partially.

- With 5 leave nodes, the number of possible rooted binary trees equals 105.
- The pulley principle tells us only to consider the unrooted binary trees, a total of 15.

Likelihood maximization

Likelihood maximization

To maximize the log-likelihood:

- Step 1: Select a tree topology.
- Step 2: Choose initial values for each edge.
- Step 3: Maximize edges individually, given the other edges.
- Step 4: Iterate step 3, until values no longer change.
- Step 5: Do this for all possible topologies.

The particular form of this algorithm described below may converge to local maxima!

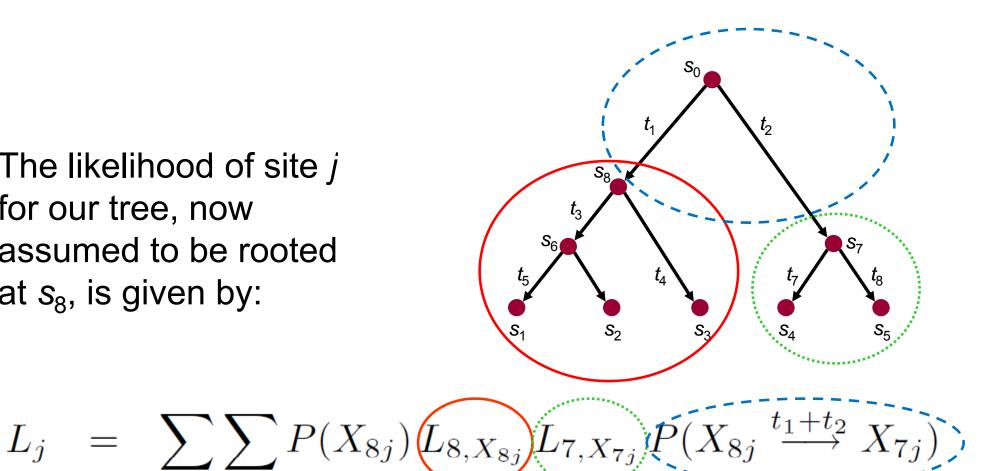
With respect to step 3: How to maximize the log-likelihood with respect to an edge?

The likelihood

Denote the conditional likelihood of subtree rooted at node *i* with nucleotide X_{ij} by $L_{i,X_{ij}}$.

The likelihood of site *j* for our tree, now assumed to be rooted at s_8 , is given by:

 $X_{8i} X_{7i}$



Likelihood maximization

Using:

$$P(X_{8j} \xrightarrow{t} X_{7j}) = \frac{1}{4} [1 - (1 - 4u)^t] + \delta_{X_{8j}, X_{7j}} (1 - 4u)^t \\ = \frac{1}{4} (1 - p) + \delta_{X_{8j}, X_{7j}} p$$

reformulate this to:

$$L_{j} = p \sum_{X_{8j}} \sum_{X_{7j}} \delta_{X_{8j}, X_{7j}} P(X_{8j}) L_{8, X_{8j}} L_{7, X_{7j}}$$
$$+ (1 - p) \sum_{X_{8j}} \sum_{X_{7j}} \frac{1}{4} P(X_{8j}) L_{8, X_{8j}} L_{7, X_{7j}}$$
$$= A_{j} p + B_{j} (1 - p)$$

This holds for all sites, thus:

$$L = \prod_{j=1}^{25} [A_j \, p + B_j \, (1-p)]$$

The log-likelihood and its derivative are given by:

$$\log(L) = \sum_{j=1}^{25} \log[A_j p + B_j (1-p)]$$

$$\frac{\partial \log(L)}{\partial p} = \sum_{j=1}^{25} \frac{A_j - B_j}{A_j p + B_j (1-p)} = 0$$

Likelihood maximization

The p maximizing the log-likelihood is found iteratively. • Choose a step size h > 0.

- Let $p^{(k)}$ be the value of p from the k-th iteration.
- Then, define:

$$p^{(k+1)} = p^{(k)} + \frac{h}{m} \sum_{j=1}^{25} \left[\frac{A_j p^{(k)}}{A_j p^{(k)} + B_j (1 - p^{(k)})} - p^{(k)} \right]$$

This choice of $p^{(k+1)}$ implies the majorization: $\sum_{j=1}^{25} \log[A_j p^{(k+1)} + B_j (1 - p^{(k+1)})]$ $\geq \sum_{j=1}^{25} \log[A_j p^{(k)} + B_j (1 - p^{(k)})]$ The majorization can be seen from:

$$p^{(k+1)} - p^{(k)} = h p^{(k)} \left(\frac{1}{m} \sum_{j=1}^{25} \frac{A_j}{A_j p^{(k)} + B_j (1 - p^{(k)})} - 1 \right)$$
$$= h p^{(k)} \left(1 - p^{(k)} \right) \frac{1}{m} \sum_{j=1}^{25} \frac{A_j - B_j}{A_j p + B_j (1 - p)}$$
$$= h p^{(k)} \left(1 - p^{(k)} \right) \frac{1}{m} \left. \frac{\partial \log(L)}{\partial p} \right|_{p=p^{(k)}}$$

which has the same sign at the derivative of the log-likelihood, evaluated in the current estimate of *p*!

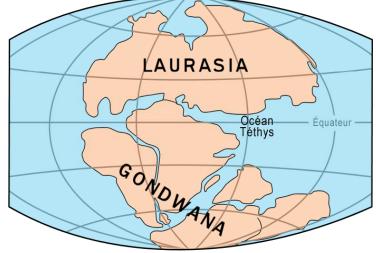
Why the likelihood approach?

Why use the likelihood approach when also the methodologically simpler *distance matrix* and *maximum parsimony methods* are available?

- The likelihood approach makes assumptions explicit. This enables us to assess their validity.
- Within the likelihood framework we may compare nested models using a likelihood ratio test.

Laurasiatheria is a group of mammals originating from the former continent Laurasia.

The phylogenetic relationships between the Laurasiatherians are still uncertain.



Available:

- RNA sequence data of 47 Laurasiatherians.
- Sequence is 3179 bases long.

Reconstruct their phylogenetic tree.



In R:

- > # activate library
- > library(phangorn)
- > # load data

. . .

> data(Laurasiatherian)

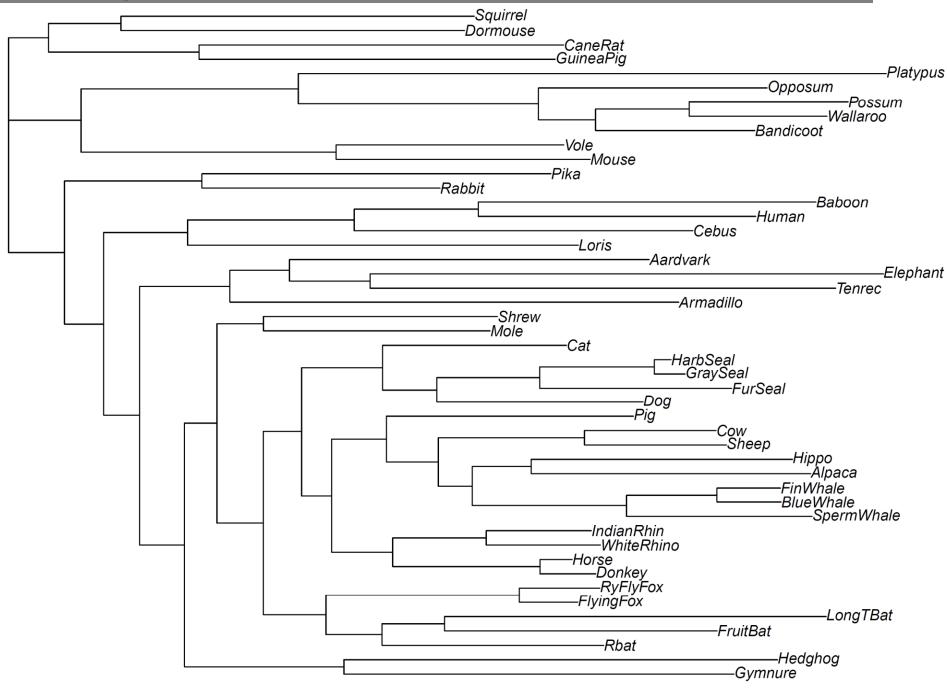
Platypus ttaaaggtttggtcctagccttactgttagatttgattagatttatacatgcagtatcc... Wallaroo ccaaaggtttggtcctggccttactgttaattgtagttagacctacacatgcagtttcc... Possum ccaaaggtttggtcctagccttactgttaattataattaaacctacacatgcagtttcc... Bandicoot ccaaaggtttggtcctagcctttctattaatttaattaaacctacacatgcagtctcc... Opposum ccataggtttggtcctagccttattattagttctaattagacctacacatgcagtttcc... Armadillo ccacaggtctggtcctagccttactattaattcataacaaaattacacatgcagtatca... Elephant ccaaaggtttggtcccggccttcttattggttactaggaaacttatacatgcagtatcc... Aardvark ttaaaggtttggtcctagcctttctattagttgacagtaaatttatacatgcagtatct... Tenrec ttaaaggtttggttctagcctttttattagttcttaataaaattatacatgcagtatcc... Hedghog aataaggtctggtcccagccttcctattttctattagtagaattacacatgcagtatca...



Now fit the model:

- > # construct a starting tree
- > distMat <- dist.logDet(Laurasiatherian)</pre>
- > tree <- NJ(distMat)</pre>

Note: this fits a model with continuous time, instead of discrete time as treated in the lecture.



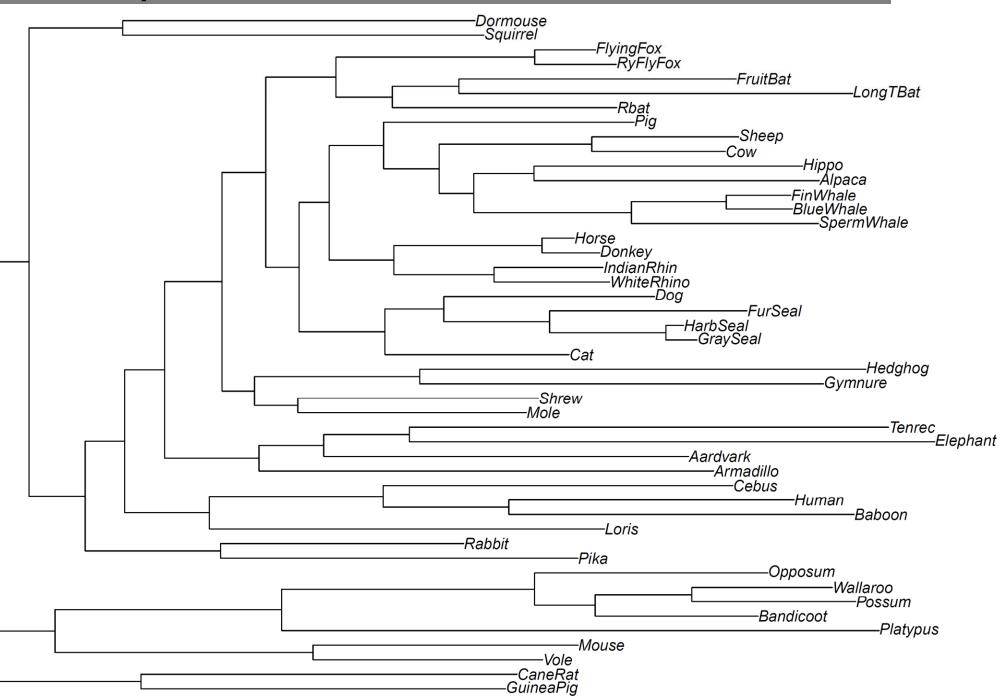


The Jukes-Cantor model is just one evolutionary model. Many more exist.

Fit different model:

- > # construct a starting tree
- > distMat <- dist.logDet(Laurasiatherian)</pre>
- > tree <- NJ(distMat)</pre>

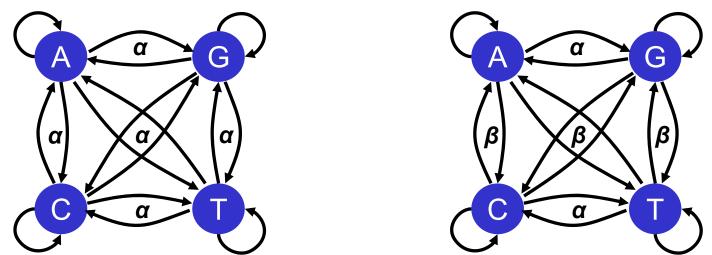
> plot(fitF81\$tree)



Transition-substitution: A, G \rightarrow A, G C, T \rightarrow C, T

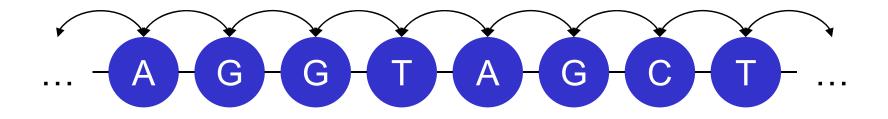
Transversion-substitution: A, G \rightarrow C, T C, T \rightarrow A, G

Transition- and transversion probabilities differ:



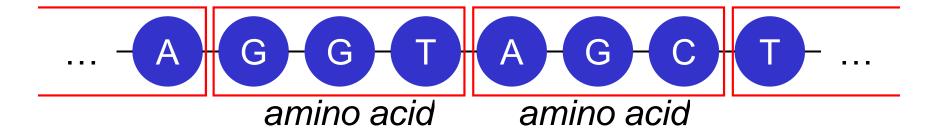
More complicated models than Jukes-Cantor available.

Positions do not evolve independently (covarion):



But also ...

... three contiguous bases code for one amino acid:



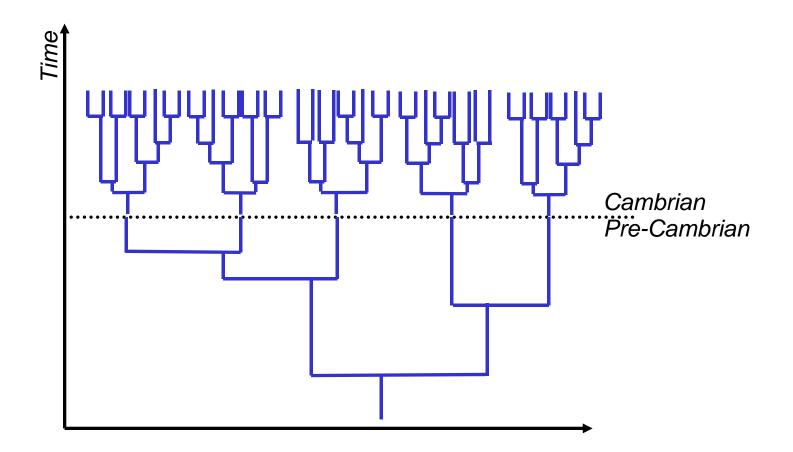
Heterotachy is a general term for within-site rate variation over time. Under heterotachy, evolutionary rates at different sites may vary in different ways over subtrees.

Hence, under heterotachy, the time-homogeneity assumption may be invalid. That is, the rate of nucleotide substution (the transition probability) may not be constant of time.

The molecular hypothesis should be applied with care.

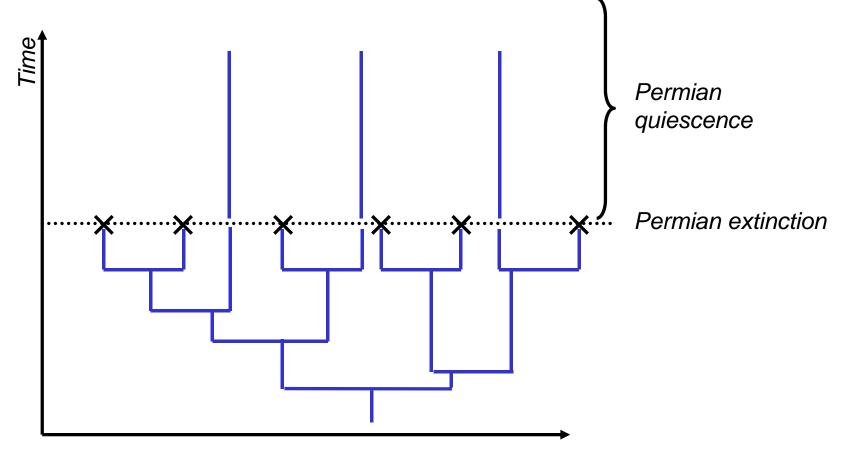
The *Cambrian explosion* refers to the period around 530 My ago in which the evolutionary pace seems accelarated.

 \rightarrow substitution-rate varies over time.



The *Permian quiescence* refers to the period after the Permian extinction (250 My ago), where the evolutionary pace seemed to have slowed down.

 \rightarrow substitution-rate varies over time.



Implicitly, it has been assumed that organism evolve independently.

However, often there is **co-evolution**:





References & further reading

References and further reading

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